Basic immunofluorescence of resin sections

Materials

Phosphate buffered saline, pH 7.4 (PBS)
Bovine serum albumen, fraction V (BSA)
Tween 20
Primary and secondary antibodies
Humid chamber

Method

1. Dry 1µm sections onto poly-l-lysine coated slides using a 37^0C hot plate.
2. Rinse with PBS-T pH 7.4 (0.1% v/v Tween 20)
3. Add blocking solution (5% (w/v) BSA in PBS-T). Incubate for 30-60min at room temperature in a humid chamber.
4. Add primary antibody, diluted in 1% BSA in PBS-T (the dilution required is very variable, from 1 in 5 to 1 in 100). Incubate for 2hrs at room temperature or overnight at 4^oC in a humid chamber
5. Wash sections with PBS-T, with several changes over 20min.
6. Add secondary antibody (with fluorescent tag), diluted in 1% BSA in PBS-T T (the dilution required is again very variable, from 1 in 100 to 1 in 1000). Incubate in the dark for 1hr at room temperature in a humid chamber.
7. Wash slides 2x with PBS-T, 2x with PBS and 3x with dh_2O.
8. Observe under fluorescence.